

1   **Caloric restriction and exercise “mimetics”: ready for prime time?**

2

3   Christoph Handschin<sup>1\*</sup>

4

5   <sup>1</sup>Biozentrum, University of Basel, Klingelbergstrasse 50/70, CH-4056 Basel, Switzerland

6

7

8

9   Published in Pharmacol Res. 2016 Jan; 103:158–66. PMID: 26658171. doi:  
10 10.1016/j.phrs.2015.11.009

11 Copyright © Elsevier; Pharmacological Research

12

13

14

15

16

17

18

19

20

### **Reprint request**

Unfortunately, due to copyright-relatedt issues, we are not able to post the post-print pdf version of our manuscript - in some cases, not even any version of our manuscript. Thus, if you would like to request a post-production, publisher pdf reprint, please click send an email with the request to christoph-dot-handschin\_at\_unibas-dot-ch (see <http://www.biozentrum.unibas.ch/handschin>).

Information about the Open Access policy of different publishers/journals can be found on the SHERPA/ROMEO webpage: <http://www.sherpa.ac.uk/romeo/>

### **Reprint Anfragen**

Aufgrund fehlender Copyright-Rechte ist es leider nicht möglich, dieses Manuskript in der finalen Version, z.T. sogar in irgendeiner Form frei zugänglich zu machen. Anfragen für Reprints per Email an christoph-dot-handschin\_at\_unibas-dot-ch (s. <http://www.biozentrum.unibas.ch/handschin>).

Informationen zur Open Access Handhabung verschiedener Verlage/Journals sind auf der SHERPA/ROMEO Webpage verfügbar: <http://www.sherpa.ac.uk/romeo/>

1   **Caloric restriction and exercise “mimetics”: ready for prime time?**

2

3   Christoph Handschin<sup>1\*</sup>

4

5   <sup>1</sup>Biozentrum, University of Basel, Klingelbergstrasse 50/70, CH-4056 Basel, Switzerland

6

7   \*Correspondence: christoph.handschin@unibas.ch

8

9

10 **Abstract**

11

12 Exercise and diet are powerful interventions to prevent and ameliorate various pathologies. The  
13 development of pharmacological agents that confer exercise- or caloric restriction-like phenotypic  
14 effects is thus an appealing therapeutic strategy in diseases or even when used as life-style and  
15 longevity drugs. Such so-called exercise or caloric restriction “mimetics” have so far mostly been  
16 described in pre-clinical, experimental settings with limited translation into humans. Interestingly,  
17 many of these compounds activate related signaling pathways, most often postulated to act on the  
18 common downstream effector peroxisome proliferator-activated receptor  $\gamma$  coactivator 1 $\alpha$  (PGC-1 $\alpha$ )  
19 in skeletal muscle. In this review, resveratrol and other exercise- and caloric restriction “mimetics”  
20 are discussed with a special focus on feasibility, chances and limitations of using such compounds in  
21 patients as well as in healthy individuals.

22

23

24

25

26

27 **Keywords:** skeletal muscle; exercise; mimetics; resveratrol; PGC-1 $\alpha$ ; AMPK; PPAR $\beta/\delta$ ; caloric  
28 restriction; diet

29

30 **Chemical compounds discussed in this article:** AICAR (PubChem CID 266934); Celastrol (PubChem  
31 CID 122724); (-)-Epicatechin (PubChem CID 72276); GSK4716 (PubChem CID 5399376); GW1516  
32 (PubChem ID 9803963); Metformin (PubChem CID 4091); Nicotinamide riboside (PubChem CID  
33 439924); Rapamycin (PubChem CID 5040); Resveratrol (PubChem CID 445154); SR9009 (PubChem ID  
34 57394020); SRT1720 (PubChem ID 25232708); Ursolic acid (PubChem CID 64945)

35

36 **Abbreviations:** ActRIIB, activin receptor type IIB; AICAR, 5-Aminoimidazole-4-carboxamide  
37 ribonucleotide; AMPK, AMP-activated protein kinase; BAIBA,  $\beta$ -aminoisobutyric acid; ERR $\gamma$ , estrogen-  
38 related receptor  $\gamma$ ; FGF21, fibroblast growth factor 21; HSF1, heat shock factor 1; MOTS-c,  
39 mitochondrial open reading frame of the 12S rRNA-c; mTOR, mammalian target of rapamycin; PGC-  
40 1 $\alpha$ , peroxisome proliferator-activated receptor  $\gamma$  coactivator 1 $\alpha$ ; PPAR $\beta/\delta$ , peroxisome proliferator-  
41 activated receptor  $\beta/\delta$ ; SARM, selective androgen receptor modulator; SIRT1, sirtuin 1

42

43

44    **1. Introduction**

45

46    The increasingly sedentary life-style and a historically unprecedented access to excess high caloric  
47    food in Western societies strongly drive an epidemic rise in many pathologies, including obesity, cardiovascular disorders, metabolic syndrome, and other chronic diseases [1]. Surprisingly, physical  
48    inactivity is also associated with a higher risk for other diseases that lack an obvious link to skeletal  
49    muscle, such as certain types of cancer, neurodegeneration and mood disorders [2]. The incidence  
50    rates of most of these chronic diseases are further exacerbated by the dramatic increase in life  
51    expectancy in developed countries [3]. Likewise, old age is strongly linked to sarcopenia, muscle  
52    wasting in aging. With an increasing geriatric population, a pathological threshold for sarcopenia-  
53    associated problems is reached in more and more individuals, in particular in those not engaging in  
54    regular physical activity [4]. Collectively, the lifestyle and expectancy in Western societies thereby  
55    result in an enormous burden on health care systems and costs [5].

56  
57    Importantly, in addition to the older segments of the population, lifestyle-associated diseases are  
58    also on the rise in young individuals at an alarming rate [6]: for example, type 2 diabetes, classically  
59    referred to as adult-onset diabetes, is now much more commonly diagnosed in children compared to  
60    type 1, so-called juvenile diabetes. This development is closely correlated to increased rates of  
61    childhood obesity and hypertension. While it is difficult to extrapolate the impact of this increased  
62    incidence of chronic diseases in children and young adults, it has been speculated that this wave in  
63    childhood obesity could lead to a slowdown or even a decline in life expectancy in Western societies  
64    [7].

65

66    **1.1. Treatment of life style-associated chronic diseases: pharmacology, caloric restriction and**  
67    **exercise**

68    From an economic point of view, lifestyle-associated pathologies are extremely attractive for drug  
69    development: these diseases affect a huge number of patients, and would necessitate a prolonged  
70    intake of pharmacological agents for prevention and treatment over years and decades. It thus  
71    seems surprising that for many of these diseases, only a handful of drugs are available, often with  
72    limited efficacy in monotherapies. For example, weight loss triggered by the four obesity drugs that  
73    are currently approved by the FDA is limited and plateaus after prolonged application [8]. In contrast,  
74    lifestyle-based interventions combining exercise and diet have an enormous potential to prevent and  
75    treat numerous chronic pathologies, in some cases even rivaling or surpassing the efficacy of drugs  
76    [9,10]. Since exercise triggers many different plastic changes in skeletal muscle and beyond [11,12],  
77    the mechanisms underlying the therapeutic effect of physical activity remain mysterious.  
78    Nevertheless, physical activity increases healthspan and life expectancy in humans, at least in  
79    epidemiological correlations [13,14]. Dietary measures have varied over the last decades, shifting the  
80    focus from lipids to proteins to carbohydrates and back. The most consistent effects however are  
81    based on a general moderate reduction of caloric intake in a well-balanced diet. A more extreme  
82    form, caloric restriction, has even been reported as one of the most powerful methods to prevent  
83    age-related diseases and improve longevity in different organisms ranging from yeast, the worm *C.*  
84    *elegans*, the fruit fly *D. melanogaster* to rodents and potentially primates [15]. Intriguingly, exercise  
85    and caloric restriction result in an overlapping phenotypic outcome e.g. in terms of mitochondrial

86 function and oxidative metabolism, reduction of reactive oxygen species, DNA stability or autophagy  
87 even though energy metabolism is affected in a diametrically opposite manner and the outcome on  
88 muscle function and body weight differ dramatically (Fig. 1). However, while the health benefits of  
89 exercise are widely accepted [14,16,17], the effects of caloric restriction are under debate. At least in  
90 certain settings, caloric restriction fails to affect lifespan, or might even have a negative effect, for  
91 example in different mouse strains and different types of diets [18,19]. Reduced caloric intake often  
92 is associated with a dormant stage accompanied by reduced fertility and reproduction, e.g. spores in  
93 bacteria and fungi, Dauer larvae in *C. elegans* or torpor in mice [20]. All of these processes are of little  
94 physiological relevance in humans where reduced fertility mostly occurs with starvation and is likely  
95 uncoupled from longevity. Moreover, while health benefits have been observed upon caloric  
96 restriction in studies in rodents and primates, it is conceivable that the relative amelioration by  
97 caloric restriction is at least in part due to the metabolic deterioration in the *ad libitum* fed control  
98 groups [19]. In particular in caloric restriction studies in rhesus monkeys, this confounding aspect  
99 might have contributed to the somewhat conflicting results [21,22]. Thus, the outcome of caloric  
100 restriction on human health and life expectancy is difficult to extrapolate at the moment and variants  
101 of this approach in the form of intermittent fasting [23] or even time-restricted feeding without an  
102 overall reduction in caloric intake [24] are being tested.

103 Even though exercise and diet have been strongly linked to the prevention and treatment of different  
104 chronic diseases, compliance levels for both interventions in patients and healthy individuals are low.  
105 Caloric restriction studies often use a 30% reduction in caloric intake to achieve health benefits in  
106 animal studies. In humans, it is not clear what the baseline in caloric intake should be; regardless, a  
107 reduction of 30% would constitute a massive intervention. Exercise regimes are hampered by poor  
108 physical conditions (e.g. obesity), lack of time and motivation, depression as well as other factors  
109 [25]. Moreover, some patients are exercise intolerant, e.g. those suffering from chronic heart failure  
110 [26]. Thus, to overcome limitations in the application of caloric restriction and exercise in patients,  
111 pharmacological approaches to elicit the beneficial effects of these two interventions have been  
112 proposed in the form of caloric restriction and exercise “mimetics” [15,27].

113

## 114 **2. Caloric restriction and exercise “mimetics”**

115

116 The concept of designing pharmacological agents that engage the same or at least similar biological  
117 programs as *bona fide* training was initially focused on facultative energy expenditure [28]. Later  
118 definitions aimed at a broader effect, often with the main endpoint of increased endurance capacity  
119 [29]. Various compounds have in the meantime been tested in animal models, primarily based on the  
120 current knowledge about signaling pathways in exercise adaptation in skeletal muscle [27].  
121 Intriguingly, since at least some of these pathways are also engaged in caloric restriction, several  
122 compounds could constitute both exercise as well as caloric restriction “mimetics”. However, in the  
123 latter case, inhibitors of anabolic pathways, in particular of the mammalian target of rapamycin  
124 (mTOR) kinase pathway, seem to show most promise in terms of longevity. In animal models, also  
125 other signaling pathways involved in nutrient sensing such as the insulin-, insulin-like growth factor 1  
126 or growth hormone-triggered cascades were associated with modulations in lifespan [20,30]. Some  
127 examples for both classes of “mimetics” will be discussed in the following sections.

128

129 **2.1. Exercise “mimetics”**

130

131 Many substances for performance enhancement exist and are widely and illegally used as doping in  
132 sports, including steroids and other anabolic hormones such as growth hormone or insulin-like  
133 growth factor 1, or  $\beta$ 2 adrenoreceptor agonists [31,32]. However, most of these compounds have  
134 limited effects in the treatment of diseases. Moreover, when used in non-replacement therapies (as  
135 in sports), steroids, growth hormones and  $\beta$ 2 agonists can elicit massive, in some cases life-  
136 threatening side-effects. Therefore, alternative ways of mimicking exercise have to be investigated.  
137 Indeed, various other molecules have been linked to improved muscle function, endurance capacity,  
138 muscle mass and strength in experimental settings. For most of these putative exercise “mimetics”,  
139 mechanisms of action have been proposed: interestingly, those compounds that lead to an  
140 improvement in muscle endurance almost always activate signaling pathways centered on the  
141 peroxisome proliferator-activated receptor  $\gamma$  coactivator 1 $\alpha$  (PGC-1 $\alpha$ ) (Fig. 2). This transcriptional  
142 coregulator is one of the regulatory nexi of the endurance phenotype in muscle [12,33]. Accordingly,  
143 muscle-specific PGC-1 $\alpha$  transgenic mice exhibit a shift towards oxidative, high endurance muscle  
144 fibers, increased oxidative metabolism as well as improved fatigue resistance and endurance capacity  
145 [34]. Inversely, signs of pathological inactivity are observed in muscle-specific PGC-1 $\alpha$  knockout  
146 animals, including local and systemic inflammation and activity-induced fiber damage [35,36].  
147 Importantly however, mice with a muscle-specific ablation of PGC-1 $\alpha$  can at least in part still adapt to  
148 training indicating that alternative mechanisms can compensate for the absence of PGC-1 $\alpha$  [33].  
149 Some exercise “mimetics” could likewise circumvent muscle PGC-1 $\alpha$  in specific contexts, e.g. as  
150 shown for resveratrol [37].

151

152 **2.1.1. Resveratrol and SRT1720**

153

154 Resveratrol is a naturally occurring polyphenol primarily found in plants [38]. Resveratrol has  
155 pleiotropic properties and can act as an anti-inflammatory and antioxidant molecule, a  
156 phytoestrogen, an activator of the AMP-activated protein kinase (AMPK) and the ataxia-  
157 telangiectasia mutated kinase, an inhibitor of phosphodiesterases, of the F1-ATPase and of  
158 cyclooxygenase 1 as well as a modulator of the activity of complex I of the respiratory chain [38].  
159 Most prominently, resveratrol has been postulated as a direct activator of sirtuin 1 (SIRT1) [39] –  
160 however, this model of direct activation has been challenged [40] and alternative pathways of  
161 resveratrol-dependent indirect activation of SIRT1 proposed, e.g. through AMPK [41]. Similarly, the  
162 direct activation of other pharmacological SIRT1 modulators [42] has likewise been questioned, for  
163 example that of SRT1720 [40]. Direct and indirect activation of SIRT1 results in protein deacetylation  
164 of PGC-1 $\alpha$ , which is associated with higher transcriptional activity of this coactivator [43]. Moreover,  
165 the boost in cAMP levels upon resveratrol-mediated inhibition of phosphodiesterases not only  
166 directly activates SIRT1 in an NAD $^+$ -independent manner [44], but also promotes transcription of the  
167 PGC-1 $\alpha$  gene [45]. Thus, by engaging multiple signaling pathways, both resveratrol and SRT1720  
168 were shown to improve oxidative metabolism and endurance capacity in mice [46,47]. Of note, many

169 of the beneficial effects of resveratrol and SRT1720 on systemic metabolic parameters also occur in  
170 muscle-specific PGC-1 $\alpha$  knockout animals indicating redundant signaling pathways and/or  
171 engagement of PGC-1 $\alpha$  and other targets in non-muscle tissue to mediate these systemic effects  
172 [37]. Importantly, these observations were only reported in rodents fed a high fat-containing diet,  
173 which most likely suffer from impaired muscle endurance: it thus is unclear whether similar effects  
174 on exercise capacity would also be observed in mice on a regular diet. Moreover, the reported effect  
175 of resveratrol on mitochondrial biogenesis in skeletal muscle was not replicated in all studies to the  
176 same extent [37,48]. Then, in mice, resveratrol and SRT1720 exhibit an organ preference for the  
177 modulation of metabolic pathways by primarily affecting white adipose tissue and liver, respectively,  
178 implying different tissue bioavailability and/or molecular targets for these two compounds [37].  
179 Finally, and most alarmingly, resveratrol application in human exercise studies surprisingly blunted  
180 many aspects of training adaptation [49-51]. Even though the pathways mediating this unexpected  
181 detrimental effect of resveratrol on training in humans are not clear, similar findings have previously  
182 been reported for other antioxidants [52]. Thus, more studies are needed to resolve the molecular  
183 mechanisms and reveal whether resveratrol and SRT1720 really promote exercise-like effects in  
184 skeletal muscle of healthy mice and humans. Other aspects of resveratrol application are discussed in  
185 section 2.2.1.

186

### 187 **2.1.2. AMPK activators (e.g. AICAR and metformin)**

188

189 5-Aminoimidazole-4-carboxamide ribonucleotide (AICAR) is an intermediate metabolite of the  
190 inosine monophosphate biosynthesis pathway. As an analog of AMP, AICAR activates AMPK. This  
191 kinase is centrally involved in the response of skeletal muscle fibers to contraction and exercise [53].  
192 Accordingly, pharmacological activation of AMPK triggers many of the posttranslational and  
193 transcriptional adaptations to endurance training, collectively resulting in mice with a higher  
194 endurance capacity [29]. At least in part, this response is due to AMPK-dependent phosphorylation  
195 and transcriptional activation of the PGC-1 $\alpha$  protein and gene, respectively [54]. Metformin, one of  
196 the most widely used drugs in the treatment of type 2 diabetes, also activates AMPK [55]. However, a  
197 slight reduction in exercise-related parameters including VO<sub>2max</sub> or exercise duration in healthy  
198 individuals and a blunting of the effects of training in prediabetic individuals have been reported [27].  
199 These negative effects of metformin on exercise parameters could stem from the metformin-  
200 dependent inhibition of complex I of mitochondrial oxidative phosphorylation, which by itself is  
201 sufficient to decreases exercise performance [27]. Therefore, non-selective activators of AMPK  
202 should be considered with caution as exercise “mimetics”. Moreover, the use of AICAR and other  
203 specific AMPK modulators might be hampered by poor bioavailability as well as the effect of long-  
204 term activation of AMPK to increase circulating levels of lactic and uric acid and promote a chronic  
205 catabolic state by inhibiting mTOR signaling [56].

206

### 207 **2.1.3. PPAR $\beta/\delta$ activators (e.g. GW1516)**

208

209 The levels of the peroxisome proliferator-activated receptor  $\beta/\delta$  (PPAR $\beta/\delta$ , NR1C2) in skeletal muscle  
210 are regulated by exercise and in turn, this nuclear receptor promotes the expression of genes  
211 encoding proteins involved in fatty acid oxidation and mitochondrial substrate metabolism [57]. This  
212 transcriptional control of exercise genes could be mediated by PPAR $\beta/\delta$  control of PGC-1 $\alpha$  expression  
213 [57]. Inversely, PGC-1 $\alpha$  can exert its potent effects on muscle transcription in the absence of  
214 PPAR $\beta/\delta$  [58] even though PGC-1 $\alpha$  coactivates PPAR $\beta/\delta$  in skeletal muscle [59]. GW1516, a synthetic  
215 ligand of PPAR $\beta/\delta$ , improves fatty acid oxidation in skeletal muscle. Moreover, while GW1516 is  
216 insufficient to improve endurance in untrained mice, pharmacological activation of PPAR $\beta/\delta$  boosts  
217 the effects of endurance exercise [29]. Importantly, concomitant application of AICAR and GW1516  
218 potentiates the effects of each individual compound on mitochondrial gene transcription and  
219 endurance capacity [29]. Therefore, activation of PPAR $\beta/\delta$  might be an option to further improve the  
220 effect of exercise or of other exercise "mimetics". However, open questions about potential pro-  
221 tumorigenic effects of PPAR $\beta/\delta$  will have to be resolved before such activators can be used in  
222 humans [60].

223

#### 224 **2.1.4. ERR $\gamma$ agonists (e.g. GSK4716)**

225

226 The nuclear receptor estrogen-related receptor  $\gamma$  (ERR $\gamma$ , NR3B3) is a potent regulator of oxidative  
227 metabolism in skeletal muscle and other tissues. GSK4716, a synthetic ligand of ERR $\gamma$ , promotes an  
228 endurance-trained phenotype in mice concomitant with enhanced mitochondrial function,  
229 vascularization and a switch towards slow, oxidative muscle fibers [61]. Mechanistically,  
230 overexpression of ERR $\gamma$  in skeletal muscle in mice results in activation of AMPK and, somewhat  
231 surprisingly, no change in PGC-1 $\alpha$  transcript levels or protein acetylation [61]. Intriguingly however,  
232 short-term treatment of primary muscle cells with GSK4716 leads to a robust induction of PGC-1 $\alpha$   
233 gene transcription [62] revealing a discrepancy between acute and chronic activation of ERR $\gamma$  in  
234 ligand-treated muscle cells and transgenic animals, respectively. Once both proteins are activated,  
235 ERR $\gamma$  and PGC-1 $\alpha$  proteins interact to form a transcriptionally active complex in the regulation of  
236 metabolic target genes [63].

237

#### 238 **2.1.5. REV-ERB $\alpha$ agonists (e.g. SR9009)**

239

240 REV-ERB $\alpha$  (NR1D1) is a nuclear receptor with a dual role in the control of circadian rhythm and of  
241 metabolism [64]. In skeletal muscle, REV-ERB $\alpha$  activity controls mitochondrial biogenesis, autophagy  
242 and other processes that culminate in a higher endurance capacity in mice [65]. Mechanistically,  
243 muscle-specific ablation of REV-ERB $\alpha$  results in reduced activity of the AMPK-SIRT1-PGC-1 $\alpha$  signaling  
244 pathway. Inversely, these regulators of exercise adaptation are activated in muscle-specific REV-ERB $\alpha$   
245 overexpressing mice or animals treated with SR9009, a synthetic agonist of REV-ERB $\alpha$  [65].  
246 Moreover, PGC-1 $\alpha$  controls REV-ERB $\alpha$  expression by coactivating the retinoic acid receptor-related  
247 orphan receptor  $\alpha$  (NR1F1) [66]. Interestingly, application of SR9009 does not affect the muscle fiber-  
248 type distribution. Moreover, the effects of SR9009 on the circadian functions of REV-ERB $\alpha$  lead to a  
249 change in circadian behavior and circadian rhythm core regulatory genes in the hypothalamus [67].

250 The circadian expression pattern of various genes in the liver, skeletal muscle and fat is likewise  
251 affected [67]. Thus, the consequences of using SR9009 as an exercise “mimetic” in terms of circadian  
252 rhythms in humans will have to be carefully evaluated.

253

254 **2.1.6. Resistance training “mimetics” (e.g. myostatin pathway inhibitors, selective androgen  
255 receptor modulators and ursolic acid)**

256

257 Surprisingly, almost all of the currently described experimental exercise “mimetics” promote an  
258 endurance-trained state, in comparison to the relatively few candidates that would trigger a  
259 resistance training-like phenotype. At least in part, this might be due to the well-established  
260 endurance exercise training protocols using running wheels, treadmills or swimming as paradigms in  
261 contrast to the limited possibilities to perform resistance training in rodents. Accordingly, the design  
262 and application of the currently most promising candidates, myostatin pathway inhibitors and  
263 selective androgen receptor modulators (SARMs), were to a large extent based on data from other  
264 species. For example, naturally occurring mutations in the myostatin gene have been associated with  
265 exacerbated muscle mass in cattle and dog strains, which later could be replicated in transgenic mice  
266 [68]. However, several approaches to directly inhibit myostatin in human trials exhibited low clinical  
267 efficacy, potentially due to redundant signaling through the receptor for myostatin, the activin  
268 receptor type IIB (ActRIIB) [69]. Indeed, inhibitors of ActRIIB activation had a high therapeutic  
269 potential in animal models [70] and various forms are currently being tested in clinical trials [71].  
270 Similarly, SARMs are also being tested in clinical trials to prevent muscle wasting in different settings  
271 [72]. Steroidal and non-steroidal SARMs were designed to achieve partial activation of the androgen  
272 receptor (NR3C4), the main receptor for the androgenic steroids testosterone and  
273 dihydrotestosterone [73]. Androgenic steroids have potent anabolic effects on muscle tissue, but  
274 also exert androgenic actions, e.g. on the prostate gland. In order to act therapeutically while  
275 circumventing unwanted effects, SARMs stimulate the anabolic, but not the androgenic downstream  
276 programs controlled by the androgen receptor [72,73].

277 Ursolic acid was identified in a screen comparing the gene expression pattern of different human  
278 settings of muscle atrophy with those of the Connectivity map, a collection of global gene expression  
279 data of various compounds in different cell types [74]. In mice, ursolic acid not only blunts muscle  
280 atrophy in disease contexts, but also enhances muscle weight, fiber size and strength in healthy  
281 rodents [74]. More studies in mice and humans will reveal the robustness of ursolic acid as a  
282 resistance training exercise “mimetic”.

283

284 **2.1.7. Other candidates (e.g. myokines, mitokines, epicatechin, celastrol)**

285

286 In addition to the compounds described above, various other agents have been linked to increased  
287 muscle function or at least a partial exercise-like effect that could potentially be exploited in patients.  
288 Skeletal muscle produces and secretes a number of auto-, para- or endocrine-acting messengers,  
289 referred to as myokines [12]. Based on their effector profiles, the use of some of these myokines

could be interesting to achieve specific therapeutic goals. For example, irisin and meteorin-like are two exercise-controlled myokines under the transcriptional control of the PGC-1 $\alpha$  transcript variants PGC-1 $\alpha$ 1 and PGC-1 $\alpha$ 4, respectively [75,76]. In turn, irisin induces PGC-1 $\alpha$  gene transcription in muscle tissue and other cell types [77]. Both irisin and meteorin-like have been linked to a browning of white adipose tissue and thereby, an increase in energy expenditure. Accordingly, injection of irisin into obese mice results in weight loss and improvement of glucose homeostasis [75]. Intriguingly, the biosynthesis of the metabolite  $\beta$ -aminoisobutyric acid (BAIBA) is likewise under control of PGC-1 $\alpha$  in skeletal muscle and also induces the formation of beige adipocytes in white adipose depots [78]. Accordingly, serum levels of BAIBA in humans rise with exercise and are inversely correlated with risk factors for metabolic diseases [78]. Finally, a 16-amino acid peptide called mitochondrial open reading frame of the 12S rRNA-c (MOTS-c) is encoded in the mitochondrial genome, and hence called a mitokine, and primarily affects skeletal muscle by indirectly activating AMPK [79]. Thereby, energy expenditure is elevated and adiposity as well as insulin sensitivity in obese mice are improved. These examples of myo- and mitokines illustrate how such agents could potentially be used as partial exercise “mimetics” to activate energy expenditure in obese or type 2 diabetic patients. Many other compounds might act similarly as summarized in a recent review by Philp and colleagues [80].

Finally, a myriad of other substances have been implicated in regulating muscle function, most of which still await replication and confirmation. For example, (-)-epicatechin and celastrol are two agents proposed to act in a mechanistically different manner compared to other endurance exercise mimetics. The flavonoid (-)-epicatechin boosts tissue vascularization by activating nitric oxide signaling resulting in higher levels of the vascular endothelial growth factor [81]. Together with an effect on mitochondrial function, (-)-epicatechin-regulated enhancement of vascularization leads to an improvement in endurance capacity in mice [81]. Of note, nitric oxide signaling is also a strong activator of PGC-1 $\alpha$  in muscle [82]. Celastrol, a plant metabolite, activates the heat shock factor 1 (HSF1) and thereby engages the cellular response to heat, cold and related stress pathways [83]. Intriguingly, celastrol promotes mitochondrial function and oxidative metabolism in skeletal muscle via an HSF1-PGC-1 $\alpha$  axis and thereby is sufficient to enhance endurance capacity, at least in high fat diet-fed mice [84]. In addition to transcriptional activation of PGC-1 $\alpha$  gene expression, the interaction of these two proteins suggests coactivation of HSF1 by PGC-1 $\alpha$  to be involved in the regulation of target gene expression [84].

321

## 322 **2.2. Caloric restriction “mimetics”**

323

Caloric restriction “mimetics” are compounds that elicit similar metabolic effects as caloric restriction, activate the corresponding stress pathways and cellular protection, and extend health- and lifespan [15,30]. In fact, the last property, longevity, often served as the primary endpoint in model organisms to identify mechanisms of caloric restriction. Not surprisingly, nutrient sensors and anabolic signaling pathways were found to be central in this process. Thus, SIRT1, AMPK and the mTOR signaling pathway belong to the main targets of caloric restriction “mimetics”, mechanistically similar to the compounds used as exercise “mimetics”. Accordingly, considerable overlap exists between the two categories.

332

333 **2.2.1. Resveratrol and nicotinamide riboside**

334

335 Resveratrol was initially described to prolong lifespan in lower organisms, an effect thought to be  
336 mediated by sirtuins [39]. In rodents, improved longevity due to resveratrol administration has been  
337 challenged and could be restricted to specific mouse strains [85]. Nevertheless, an improvement of  
338 various health parameters was observed in resveratrol-fed animals even in the absence of lifespan  
339 extension, importantly however only in mice fed a high fat diet [86]. Therefore, resveratrol  
340 administration is being tested in a number of clinical trials to improve healthspan in normal  
341 individuals, and cardiovascular, metabolic and a number of other pathologies in patients [87]. In  
342 healthy individuals, beneficial effects so far either were non-existent or minor [38,87], in exercise  
343 studies even detrimental (see Section 2.1.1). In patient studies, some small, but significant  
344 improvements were observed; however, collective interpretation of the results is hampered by small  
345 study size as well as differences in doses and application [38,87]. Thus, more studies are needed to  
346 resolve open questions about mechanisms, bioavailability, toxicity, dose and interactions [88,89].

347 Besides pharmacological means, SIRT1 activation can also be promoted by modulation of the co-  
348 substrate NAD<sup>+</sup> [90]. An increase in intracellular NAD<sup>+</sup> is for example achieved by inhibition of other  
349 NAD<sup>+</sup> consumers such as poly(ADP-ribose) polymerases or by providing precursor metabolites,  
350 including nicotinamide mononucleotide or nicotinamide riboside. The latter strategy circumvents  
351 potential side-effects of poly(ADP-ribose) polymerase inhibition on other cellular processes. Indeed,  
352 both approaches improved metabolic health in high fat-diet fed mice [90] while efficacy in humans  
353 remains largely unexplored.

354

355 **2.2.2. mTOR inhibitors (e.g. rapamycin)**

356

357 The central regulator of cell growth, mTOR, is a regulatory key step in anabolic processes, in  
358 particular protein synthesis, and accordingly, mTOR signaling is reduced in most caloric restriction  
359 studies [91]. Mechanistically, this reduction in mTOR activity stems from the absence of positive  
360 inputs via nutrients and insulin, as well as a potent inhibition by AMPK. Rapamycin, a natural  
361 compound that inhibits the activity of the mTOR complex 1 and, at higher doses and when  
362 administered chronically, also mTOR complex 2, is used clinically for immunosuppression and the  
363 treatment of certain types of cancer. Interestingly, rapamycin also exerts robust effects on longevity  
364 in various species, including genetically heterologous mice [85]. The effects of rapamycin on  
365 healthspan are more controversial, and potential side effects include dysregulation of glucose and  
366 insulin homeostasis, cataracts and obviously immunosuppression [30]. There however is evidence  
367 that these unwanted effects could be avoided with appropriate dose and timing of rapamycin  
368 administration as well as more specific mTOR complex 1 inhibitors [30]. Intriguingly, mTOR and PGC-  
369 1 $\alpha$  activities intersect in regards to the regulation of mitochondrial genes via the transcription factor  
370 ying yang 1 in muscle [92]. Whether this mTOR-PGC-1 $\alpha$  crosstalk is involved in the longevity effects  
371 of rapamycin remains unclear.

372

373 **2.2.3. Metformin**

374

375 Metformin is a biguanide drug that is widely used for the treatment of type 2 diabetes. Moreover,  
376 experimentally, metformin improves lifespan of mice, at least in some studies [93]. Which of the  
377 poly-pharmacological effects of metformin are responsible for this improvement is unclear:  
378 hypothetically, metformin-mediated activation of AMPK could result in inhibition of mTOR activity  
379 and thereby prolong survival. Curiously, the effect of metformin on lifespan differs dramatically in  
380 different species and doses of application [30]. The broad clinical usage of metformin allows  
381 epidemiological assessment of health- and survival in human patients and indeed, a beneficial effect  
382 of metformin on pathological parameters and survival has been reported in different studies even  
383 though in a recent meta-analysis, no significant overall mortality benefit was found [30].  
384 Nevertheless, large clinical trials are currently being designed to study the effect of metformin in  
385 elderly individuals [94].

386

387 **2.2.4. Other candidates (e.g. glycolysis inhibitors, mitochondrial uncouplers, fibroblast growth  
388 factor 21)**

389

390 Other strategies to elicit phenotypic effects that resemble caloric restriction have aimed at reducing  
391 the ingestion, uptake and metabolism of lipids and carbohydrates [30]. For example, inhibition of  
392 glycolysis using 2-deoxy-D-glucose elicits several cellular hallmarks of caloric restriction [30]. Due to  
393 toxic effects of 2-deoxy-D-glucose for example on cardiac tissue in rats, other inhibitors of glycolysis  
394 are currently being tested [30]. Historically, another approach was used to achieve increased energy  
395 expenditure and thereby reach a caloric restriction-like state: 2,4-dinitrophenol, a mitochondrial  
396 uncoupler, was widely used as weight loss drug in the 1930s in the United States [95]. However, due  
397 to the potential severe toxicity in terms of cataracts and lethal hyperthermia, the use of  
398 pharmacological uncouplers has been discontinued. Nevertheless, exploitation of the underlying  
399 principle in a more targeted manner is still being pursued by using endogenous regulators of  
400 mitochondrial uncoupling in brown and beige adipocytes (see Section 2.1.7). Analogous to the  
401 myokines described in this section, other endogenous hormones can also trigger systemic effects  
402 resembling caloric restriction. For example, the fibroblast growth factor 21 (FGF21) is a hormone that  
403 is primarily produced by and secreted from the liver upon starvation and in turn controls the  
404 metabolic adaptation of various tissues in the body [96]. Modulation of FGF21 has profound effects  
405 on metabolic parameters in animal models of diabetes and transgenic overexpression of FGF21  
406 extends the lifespan of mice [97]. Pharmacological and protein-analogs of FGF21 are currently being  
407 tested in different clinical trials [96].

408

409 **3. Limitations and caveats**

410

411 First, most of the “mimetics” discussed here have the capacity to improve metabolic and other  
412 parameters in pathological conditions or specific diets, but only few of these compounds affect  
413 healthy mice in a physiologically relevant manner. Accordingly, human trials, e.g. with resveratrol,  
414 resulted in a small amelioration in patients, but revealed little effects in healthy individuals.  
415 Therefore, the use of existing exercise and caloric restriction “mimetics” as prevention will have to be  
416 carefully evaluated. Second, it is unclear whether single compounds can elicit all of the complex local  
417 and systemic changes that are observed after exercise or caloric restriction. Pharmacological,  
418 physiological and economic arguments comparing drugs to training have very elegantly been  
419 described by Booth and Laye [16]. For example, exercise has an extremely high therapeutic index  
420 compared to drugs. Finally, even if the design of real exercise and caloric restriction “mimetics” was  
421 feasible, application in prevention and treatment might be hampered by unwanted effects. In many  
422 regards, muscle-specific PGC-1 $\alpha$  transgenic animals could serve as a model of a genetic exercise  
423 “mimetic” [34,98]. Despite the potent positive effects on exercise parameters, this and related  
424 mouse lines depict several limitations of inducing exercise-like effects in the absence of *bona fide*  
425 physical activity. For example, when fed a high fat-containing diet, PGC-1 $\alpha$  muscle transgenic animals  
426 exhibit an accelerated development of insulin resistance instead of the expected protection that is  
427 conferred by exercise [99]. Conceivably, this paradoxical observation is caused by the ability of PGC-  
428 1 $\alpha$  not only to promote catabolic, but also anabolic processes, including synthesis and storage of  
429 intramyocellular glycogen and lipids [100]. These adaptations are likewise expected in endurance  
430 training and underlie the so-called “athlete’s paradox”, the observation of intramyocellular lipid  
431 accumulation in endurance athletes and type 2 diabetic patients [101]. Muscle-specific  
432 overexpression of PGC-1 $\alpha$  (or a pharmacological exercise “mimetic”) in sedentary animals thus  
433 triggers lipid accumulation as part of the normal exercise response. This physiological process  
434 however is exacerbated by dietary lipids in a high fat diet and therefore promotes the development  
435 of insulin resistance [100]. In athletes, accumulation of intramyocellular lipids might not be  
436 detrimental due to the constant substrate turnover in contraction-recuperation cycles: accordingly,  
437 muscle-specific PGC-1 $\alpha$  transgenic mice on a high fat diet exhibit markedly improved insulin  
438 sensitivity with concomitant physical activity to an even higher extent compared to wild-type control  
439 animals [102]. These findings imply that in addition to the transcriptional and translational changes  
440 that are elicited by an exercise “mimetic”, other processes that are controlled by physical activity  
441 such as substrate turnover are required to achieve health benefits in certain contexts [103]. In fact,  
442 without changes in physical activity and diet, application of an exercise “mimetic” could thus be  
443 detrimental as observed in the high fat diet-fed, sedentary PGC-1 $\alpha$  muscle-specific transgenic mice.  
444 Furthermore, the amount of muscle PGC-1 $\alpha$  has to be carefully titrated to avoid unwanted effects as  
445 excessively high levels of PGC-1 $\alpha$  in cardiac and skeletal muscle result in severe pathologies in either  
446 tissue [33,104]. Finally, selectivity of pharmacological activation of exercise-controlled signaling  
447 pathways should be considered since PGC-1 $\alpha$ -mediated effects in liver and pancreas could for  
448 example outweigh the beneficial effects of muscle PGC-1 $\alpha$  in regulating systemic glucose  
449 homeostasis [104]. Thus, in many cases, partial exercise “mimetics” might be a safer and more  
450 efficacious approach to alleviate specific pathologies.

451 The same arguments in regard to exercise “mimetics” could likewise be made for caloric restriction  
452 “mimetics”. In addition, other caveats exist for this class of drugs: for example, exacerbated weight  
453 loss, alterations in the balance between mitochondrial activity, membrane potential and reactive  
454 oxygen species-production, dietary composition, the genetic background and other parameters can  
455 in certain contexts result in shortening of lifespan upon caloric restriction [18]. Moreover, at least

456 some proposed caloric restriction “mimetics” alter nutrient balance and thereby also uptake of  
457 vitamins. Finally, caloric restriction can also impair immune system function leading to delayed  
458 wound healing and higher susceptibility for infections [21].

459

460 **4. Summary and conclusion**

461

462 Based on the many health benefits of exercise and diet, the concept of developing pharmacological  
463 agents that trigger similar phenotypic effects is highly attractive. However, at the moment, it is  
464 unclear if such an approach is feasible or even desirable. More research is required to better  
465 understand the molecular mechanisms that underlie cellular plasticity as well as the systemic cross-  
466 talk between organs and tissues in exercise or caloric restriction. Second, potential unwanted effects  
467 of exercise and caloric restriction “mimetics” have to be identified and confined. Third, the strategy  
468 to design partial instead of full exercise and caloric restriction “mimetics” might be more efficient to  
469 be used as drugs in specific pathological contexts. In any case, it is unlikely that such pharmacological  
470 approaches can be used without accompanying interventions based on actual physical activity and  
471 diet – thus, the economically appealing idea of a drug to be used without changes in life-style for  
472 weight loss and improved muscle function most likely will remain elusive. Similarly, since most  
473 beneficial effects in clinical trials so far were observed in patients and not in healthy individuals,  
474 exercise “mimetics” might have a limited potential for performance enhancement in athletes in  
475 which the respective systems are already activated. In conclusion, except for patients that have to  
476 overcome exercise intolerance, for example in a muscular dystrophy, “mimetic”-based  
477 pharmacological approaches will most likely not exceed the status of an adjuvant therapy besides  
478 *bona fide* life-style changes.

479

480

481 **Acknowledgments**

482

483 I would like to thank Barbara Kupr, Svenia Schnyder and Natasha Whitehead for discussion and  
484 comments on the manuscript. Work in our laboratory is supported by the ERC Consolidator grant  
485 616830-MUSCLE\_NET, the Swiss National Science Foundation, SystemsX.ch, the Swiss Society for  
486 Research on Muscle Diseases (SSEM), the “Novartis Stiftung für medizinisch-biologische Forschung”,  
487 the University of Basel and the Biozentrum.

488

489

490 **References**

- 491
- 492 1 Booth FW, Chakravarthy MV, Gordon SE, Spangenburg EE. Waging war on physical inactivity:  
493 Using modern molecular ammunition against an ancient enemy. *J Appl Physiol* 2002;93:3-30.
- 494 2 Handschin C, Spiegelman BM. The role of exercise and pgc1alpha in inflammation and  
495 chronic disease. *Nature* 2008;454:463-469.
- 496 3 Fulop T, Larbi A, Witkowski JM, McElhaney J, Loeb M, Mitnitski A, Pawelec G. Aging, frailty  
497 and age-related diseases. *Biogerontology* 2010;11:547-563.
- 498 4 Marcell TJ. Sarcopenia: Causes, consequences, and preventions. *J Gerontol A Biol Sci Med Sci*  
499 2003;58:M911-916.
- 500 5 Booth FW, Hawley JA. The erosion of physical activity in western societies: An economic  
501 death march. *Diabetologia* 2015;58:1730-1734.
- 502 6 Lobstein T, Jackson-Leach R, Moodie ML, Hall KD, Gortmaker SL, Swinburn BA, James WP,  
503 Wang Y, McPherson K. Child and adolescent obesity: Part of a bigger picture. *Lancet*  
504 2015;385:2510-2520.
- 505 7 Olshansky SJ, Passaro DJ, Hershow RC, Layden J, Carnes BA, Brody J, Hayflick L, Butler RN,  
506 Allison DB, Ludwig DS. A potential decline in life expectancy in the united states in the 21st  
507 century. *N Engl J Med* 2005;352:1138-1145.
- 508 8 Kakkar AK, Dahiya N. Drug treatment of obesity: Current status and future prospects. *Eur J  
509 Intern Med* 2015;26:89-94.
- 510 9 Knowler WC, Barrett-Connor E, Fowler SE, Hamman RF, Lachin JM, Walker EA, Nathan DM.  
511 Reduction in the incidence of type 2 diabetes with lifestyle intervention or metformin. *N Engl  
512 J Med* 2002;346:393-403.
- 513 10 Mercken EM, Carboneau BA, Krzysik-Walker SM, de Cabo R. Of mice and men: The benefits  
514 of caloric restriction, exercise, and mimetics. *Ageing Res Rev* 2012;11:390-398.
- 515 11 Hoppeler H, Baum O, Lurman G, Mueller M. Molecular mechanisms of muscle plasticity with  
516 exercise. *Compr Physiol* 2011;1:1383-1412.
- 517 12 Schnyder S, Handschin C. Skeletal muscle as an endocrine organ: Pgc-1alpha, myokines and  
518 exercise. *Bone* 2015;80:115-125.
- 519 13 Fraser GE, Shavlik DJ. Ten years of life: Is it a matter of choice? *Arch Intern Med*  
520 2001;161:1645-1652.
- 521 14 Booth FW, Laye MJ, Roberts MD. Lifetime sedentary living accelerates some aspects of  
522 secondary aging. *J Appl Physiol* 2011;111:1497-1504.
- 523 15 Lee SH, Min KJ. Caloric restriction and its mimetics. *BMB Rep* 2013;46:181-187.
- 524 16 Booth FW, Laye MJ. Lack of adequate appreciation of physical exercise's complexities can  
525 pre-empt appropriate design and interpretation in scientific discovery. *J Physiol*  
526 2009;587:5527-5539.
- 527 17 Booth FW, Roberts CK, Laye MJ. Lack of exercise is a major cause of chronic diseases. *Compr  
528 Physiol* 2012;2:1143-1211.
- 529 18 Szafranski K, Mekhail K. The fine line between lifespan extension and shortening in response  
530 to caloric restriction. *Nucleus* 2014;5:56-65.
- 531 19 Sohal RS, Forster MJ. Caloric restriction and the aging process: A critique. *Free Radic Biol Med*  
532 2014;73:366-382.
- 533 20 Fontana L, Partridge L, Longo VD. Extending healthy life span--from yeast to humans. *Science*  
534 2010;328:321-326.
- 535 21 Mattison JA, Roth GS, Beasley TM, Tilmont EM, Handy AM, Herbert RL, Longo DL, Allison DB,  
536 Young JE, Bryant M, Barnard D, Ward WF, Qi W, Ingram DK, de Cabo R. Impact of caloric  
537 restriction on health and survival in rhesus monkeys from the nia study. *Nature*  
538 2012;489:318-321.

- 539 22 Colman RJ, Anderson RM, Johnson SC, Kastman EK, Kosmatka KJ, Beasley TM, Allison DB,  
540 Cruzen C, Simmons HA, Kemnitz JW, Weindruch R. Caloric restriction delays disease onset  
541 and mortality in rhesus monkeys. *Science* 2009;325:201-204.
- 542 23 Barnosky AR, Hoddy KK, Unterman TG, Varady KA. Intermittent fasting vs daily calorie  
543 restriction for type 2 diabetes prevention: A review of human findings. *Transl Res*  
544 2014;164:302-311.
- 545 24 Chaix A, Zarrinpar A, Miu P, Panda S. Time-restricted feeding is a preventative and  
546 therapeutic intervention against diverse nutritional challenges. *Cell Metab* 2014;20:991-  
547 1005.
- 548 25 van der Wal MH, Jaarsma T, Moser DK, Veeger NJ, van Gilst WH, van Veldhuisen DJ.  
549 Compliance in heart failure patients: The importance of knowledge and beliefs. *Eur Heart J*  
550 2006;27:434-440.
- 551 26 Okita K, Kinugawa S, Tsutsui H. Exercise intolerance in chronic heart failure--skeletal muscle  
552 dysfunction and potential therapies. *Circ J* 2013;77:293-300.
- 553 27 Li S, Laher I. Exercise pills: At the starting line. *Trends Pharmacol Sci* 2015
- 554 28 Himms-Hagen J. Exercise in a pill: Feasibility of energy expenditure targets. *Curr Drug Targets*  
555 CNS Neurol Disord 2004;3:389-409.
- 556 29 Narkar VA, Downes M, Yu RT, Embler E, Wang YX, Banayo E, Mihaylova MM, Nelson MC, Zou  
557 Y, Jugilon H, Kang H, Shaw RJ, Evans RM. Ampk and ppardelta agonists are exercise  
558 mimetics. *Cell* 2008;134:405-415.
- 559 30 Ingram DK, Roth GS. Calorie restriction mimetics: Can you have your cake and eat it, too?  
560 *Ageing Res Rev* 2015;20:46-62.
- 561 31 Tandon S, Bowers LD, Fedoruk MN. Treating the elite athlete: Anti-doping information for the  
562 health professional. *Mo Med* 2015;112:122-128.
- 563 32 de Hon O, Kuipers H, van Bottenburg M. Prevalence of doping use in elite sports: A review of  
564 numbers and methods. *Sports Med* 2015;45:57-69.
- 565 33 Kupr B, Handschin C. Complex coordination of cell plasticity by a pgc-1a-controlled  
566 transcriptional network in skeletal muscle. *Front Physiol* 2015;6:325.
- 567 34 Lin J, Wu H, Tarr PT, Zhang CY, Wu Z, Boss O, Michael LF, Puigserver P, Isotani E, Olson EN,  
568 Lowell BB, Bassel-Duby R, Spiegelman BM. Transcriptional co-activator pgc-1 alpha drives the  
569 formation of slow-twitch muscle fibres. *Nature* 2002;418:797-801.
- 570 35 Handschin C, Chin S, Li P, Liu F, Maratos-Flier E, Lebrasseur NK, Yan Z, Spiegelman BM.  
571 Skeletal muscle fiber-type switching, exercise intolerance, and myopathy in pgc-1alpha  
572 muscle-specific knock-out animals. *J Biol Chem* 2007;282:30014-30021.
- 573 36 Handschin C, Choi CS, Chin S, Kim S, Kawamori D, Kurpad AJ, Neubauer N, Hu J, Mootha VK,  
574 Kim YB, Kulkarni RN, Shulman GI, Spiegelman BM. Abnormal glucose homeostasis in skeletal  
575 muscle-specific pgc-1alpha knockout mice reveals skeletal muscle-pancreatic beta cell  
576 crosstalk. *J Clin Invest* 2007;117:3463-3474.
- 577 37 Svensson K, Schnyder S, Albert V, Cardel B, Quagliata L, Terracciano LM, Handschin C.  
578 Resveratrol and srt1720 elicit differential effects in metabolic organs and modulate systemic  
579 parameters independently of skeletal muscle peroxisome proliferator-activated receptor  
580 gamma co-activator 1alpha (pgc-1alpha). *J Biol Chem* 2015;290:16059-16076.
- 581 38 Bitterman JL, Chung JH. Metabolic effects of resveratrol: Addressing the controversies. *Cell*  
582 *Mol Life Sci* 2015;72:1473-1488.
- 583 39 Howitz KT, Bitterman KJ, Cohen HY, Lamming DW, Lavu S, Wood JG, Zipkin RE, Chung P,  
584 Kisielewski A, Zhang LL, Scherer B, Sinclair DA. Small molecule activators of sirtuins extend  
585 *Saccharomyces cerevisiae* lifespan. *Nature* 2003;425:191-196.
- 586 40 Pacholec M, Bleasdale JE, Chrunk B, Cunningham D, Flynn D, Garofalo RS, Griffith D, Griffor  
587 M, Loulakis P, Pabst B, Qiu X, Stockman B, Thanabal V, Varghese A, Ward J, Withka J, Ahn K.  
588 Srt1720, srt2183, srt1460, and resveratrol are not direct activators of sirt1. *J Biol Chem*  
589 2010;285:8340-8351.

- 590 41 Canto C, Gerhart-Hines Z, Feige JN, Lagouge M, Noriega L, Milne JC, Elliott PJ, Puigserver P,  
591 Auwerx J. Ampk regulates energy expenditure by modulating nad+ metabolism and sirt1  
592 activity. *Nature* 2009;458:1056-1060.
- 593 42 Milne JC, Lambert PD, Schenk S, Carney DP, Smith JJ, Gagne DJ, Jin L, Boss O, Perni RB, Vu CB,  
594 Bemis JE, Xie R, Disch JS, Ng PY, Nunes JJ, Lynch AV, Yang H, Galonek H, Israeliyan K, Choy W,  
595 Iflland A, Lavu S, Medvedik O, Sinclair DA, Olefsky JM, Jirousek MR, Elliott PJ, Westphal CH.  
596 Small molecule activators of sirt1 as therapeutics for the treatment of type 2 diabetes.  
597 *Nature* 2007;450:712-716.
- 598 43 Gerhart-Hines Z, Rodgers JT, Bare O, Lerin C, Kim SH, Mostoslavsky R, Alt FW, Wu Z,  
599 Puigserver P. Metabolic control of muscle mitochondrial function and fatty acid oxidation  
600 through sirt1/pgc-1alpha. *Embo J* 2007;26:1913-1923.
- 601 44 Gerhart-Hines Z, Dominy JE, Jr., Blattler SM, Jedrychowski MP, Banks AS, Lim JH, Chim H, Gygi  
602 SP, Puigserver P. The camp/pka pathway rapidly activates sirt1 to promote fatty acid  
603 oxidation independently of changes in nad(+). *Mol Cell* 2011;44:851-863.
- 604 45 Yoon JC, Puigserver P, Chen G, Donovan J, Wu Z, Rhee J, Adelmant G, Stafford J, Kahn CR,  
605 Granner DK, Newgard CB, Spiegelman BM. Control of hepatic gluconeogenesis through the  
606 transcriptional coactivator pgc-1. *Nature* 2001;413:131-138.
- 607 46 Feige JN, Lagouge M, Canto C, Strehle A, Houten SM, Milne JC, Lambert PD, Mataki C, Elliott  
608 PJ, Auwerx J. Specific sirt1 activation mimics low energy levels and protects against diet-  
609 induced metabolic disorders by enhancing fat oxidation. *Cell Metab* 2008;8:347-358.
- 610 47 Lagouge M, Argmann C, Gerhart-Hines Z, Meziane H, Lerin C, Daussin F, Messadeq N, Milne J,  
611 Lambert P, Elliott P, Geny B, Laakso M, Puigserver P, Auwerx J. Resveratrol improves  
612 mitochondrial function and protects against metabolic disease by activating sirt1 and pgc-  
613 1alpha. *Cell* 2006;127:1109-1122.
- 614 48 Higashida K, Kim SH, Jung SR, Asaka M, Holloszy JO, Han DH. Effects of resveratrol and sirt1  
615 on pgc-1alpha activity and mitochondrial biogenesis: A reevaluation. *PLoS Biol*  
616 2013;11:e1001603.
- 617 49 Scribbans TD, Ma JK, Edgett BA, Vorobej KA, Mitchell AS, Zelt JG, Simpson CA, Quadrilatero J,  
618 Gurd BJ. Resveratrol supplementation does not augment performance adaptations or fibre-  
619 type-specific responses to high-intensity interval training in humans. *Appl Physiol Nutr Metab*  
620 2014;39:1305-1313.
- 621 50 Gliemann L, Schmidt JF, Olesen J, Bienso RS, Peronard SL, Grandjean SU, Mortensen SP,  
622 Nyberg M, Bangsbo J, Pilegaard H, Hellsten Y. Resveratrol blunts the positive effects of  
623 exercise training on cardiovascular health in aged men. *J Physiol* 2013;591:5047-5059.
- 624 51 Olesen J, Gliemann L, Bienso R, Schmidt J, Hellsten Y, Pilegaard H. Exercise training, but not  
625 resveratrol, improves metabolic and inflammatory status in skeletal muscle of aged men. *J  
626 Physiol* 2014;592:1873-1886.
- 627 52 Ristow M, Zarse K, Oberbach A, Kloting N, Birringer M, Kiehnkopf M, Stumvoll M, Kahn CR,  
628 Bluher M. Antioxidants prevent health-promoting effects of physical exercise in humans. *Proc  
629 Natl Acad Sci U S A* 2009;106:8665-8670.
- 630 53 Mounier R, Theret M, Lantier L, Foretz M, Viollet B. Expanding roles for ampk in skeletal  
631 muscle plasticity. *Trends Endocrinol Metab* 2015;26:275-286.
- 632 54 Jager S, Handschin C, St-Pierre J, Spiegelman BM. Amp-activated protein kinase (ampk) action  
633 in skeletal muscle via direct phosphorylation of pgc-1alpha. *Proc Natl Acad Sci U S A*  
634 2007;104:12017-12022.
- 635 55 Zhou G, Myers R, Li Y, Chen Y, Shen X, Fenyk-Melody J, Wu M, Ventre J, Doepper T, Fujii N,  
636 Musi N, Hirshman MF, Goodyear LJ, Moller DE. Role of amp-activated protein kinase in  
637 mechanism of metformin action. *J Clin Invest* 2001;108:1167-1174.
- 638 56 Hawley JA, Holloszy JO. Exercise: It's the real thing! *Nutr Rev* 2009;67:172-178.
- 639 57 Schuler M, Ali F, Chambon C, Duteil D, Bornert JM, Tardivel A, Desvergne B, Wahli W,  
640 Chambon P, Metzger D. Pgc1alpha expression is controlled in skeletal muscles by pparbeta,  
641 whose ablation results in fiber-type switching, obesity, and type 2 diabetes. *Cell Metab*  
642 2006;4:407-414.

- 643 58 Perez-Schindler J, Svensson K, Vargas-Fernandez E, Santos G, Wahli W, Handschin C. The  
644 coactivator pgc-1alpha regulates skeletal muscle oxidative metabolism independently of the  
645 nuclear receptor pparbeta/delta in sedentary mice fed a regular chow diet. *Diabetologia*  
646 2014;57:2405-2412.
- 647 59 Dressel U, Allen TL, Pippal JB, Rohde PR, Lau P, Muscat GE. The peroxisome proliferator-  
648 activated receptor beta/delta agonist, gw501516, regulates the expression of genes involved  
649 in lipid catabolism and energy uncoupling in skeletal muscle cells. *Mol Endocrinol*  
650 2003;17:2477-2493.
- 651 60 Giordano Attianese GM, Desvergne B. Integrative and systemic approaches for evaluating  
652 pparbeta/delta (ppard) function. *Nucl Recept Signal* 2015;13:e001.
- 653 61 Narkar VA, Fan W, Downes M, Yu RT, Jonker JW, Alaynick WA, Banayo E, Karunasiri MS, Lorca  
654 S, Evans RM. Exercise and pgc-1alpha-independent synchronization of type i muscle  
655 metabolism and vasculature by errgamma. *Cell Metab* 2011;13:283-293.
- 656 62 Rangwala SM, Wang X, Calvo JA, Lindsley L, Zhang Y, Deyneko G, Beaulieu V, Gao J, Turner G,  
657 Markovits J. Estrogen-related receptor gamma is a key regulator of muscle mitochondrial  
658 activity and oxidative capacity. *J Biol Chem* 2010;285:22619-22629.
- 659 63 Devarakonda S, Gupta K, Chalmers MJ, Hunt JF, Griffin PR, Van Duyne GD, Spiegelman BM.  
660 Disorder-to-order transition underlies the structural basis for the assembly of a  
661 transcriptionally active pgc-1alpha/errgamma complex. *Proc Natl Acad Sci U S A*  
662 2011;108:18678-18683.
- 663 64 Cho H, Zhao X, Hatori M, Yu RT, Barish GD, Lam MT, Chong LW, DiTacchio L, Atkins AR, Glass  
664 CK, Liddle C, Auwerx J, Downes M, Panda S, Evans RM. Regulation of circadian behaviour and  
665 metabolism by rev-erb-alpha and rev-erb-beta. *Nature* 2012;485:123-127.
- 666 65 Woldt E, Sebti Y, Solt LA, Duhem C, Lancel S, Eeckhoute J, Hesselink MK, Paquet C, Delhaye S,  
667 Shin Y, Kamenecka TM, Schaart G, Lefebvre P, Neviere R, Burris TP, Schrauwen P, Staels B,  
668 Duez H. Rev-erb-alpha modulates skeletal muscle oxidative capacity by regulating  
669 mitochondrial biogenesis and autophagy. *Nat Med* 2013;19:1039-1046.
- 670 66 Liu C, Li S, Liu T, Borjigin J, Lin JD. Transcriptional coactivator pgc-1alpha integrates the  
671 mammalian clock and energy metabolism. *Nature* 2007;447:477-481.
- 672 67 Solt LA, Wang Y, Banerjee S, Hughes T, Kojetin DJ, Lundasen T, Shin Y, Liu J, Cameron MD,  
673 Noel R, Yoo SH, Takahashi JS, Butler AA, Kamenecka TM, Burris TP. Regulation of circadian  
674 behaviour and metabolism by synthetic rev-erb agonists. *Nature* 2012;485:62-68.
- 675 68 Cohen S, Nathan JA, Goldberg AL. Muscle wasting in disease: Molecular mechanisms and  
676 promising therapies. *Nat Rev Drug Discov* 2015;14:58-74.
- 677 69 Rodino-Klapac LR, Haidet AM, Kota J, Handy C, Kaspar BK, Mendell JR. Inhibition of myostatin  
678 with emphasis on follistatin as a therapy for muscle disease. *Muscle Nerve* 2009;39:283-296.
- 679 70 Zhou X, Wang JL, Lu J, Song Y, Kwak KS, Jiao Q, Rosenfeld R, Chen Q, Boone T, Simonet WS,  
680 Lacey DL, Goldberg AL, Han HQ. Reversal of cancer cachexia and muscle wasting by actriib  
681 antagonism leads to prolonged survival. *Cell* 2010;142:531-543.
- 682 71 Han HQ, Zhou X, Mitch WE, Goldberg AL. Myostatin/activin pathway antagonism: Molecular  
683 basis and therapeutic potential. *Int J Biochem Cell Biol* 2013;45:2333-2347.
- 684 72 McEwan IJ. Androgen receptor modulators: A marriage of chemistry and biology. *Future Med  
685 Chem* 2013;5:1109-1120.
- 686 73 Gao W. Androgen receptor as a therapeutic target. *Adv Drug Deliv Rev* 2010;62:1277-1284.
- 687 74 Kunkel SD, Suneja M, Ebert SM, Bongers KS, Fox DK, Malmberg SE, Alipour F, Shields RK,  
688 Adams CM. Mrna expression signatures of human skeletal muscle atrophy identify a natural  
689 compound that increases muscle mass. *Cell Metab* 2011;13:627-638.
- 690 75 Bostrom P, Wu J, Jedrychowski MP, Korde A, Ye L, Lo JC, Rasbach KA, Bostrom EA, Choi JH,  
691 Long JZ, Kajimura S, Zingaretti MC, Vind BF, Tu H, Cinti S, Holmlund K, Gygi SP, Spiegelman BM.  
692 A pgc1-alpha-dependent myokine that drives brown-fat-like development of white fat and  
693 thermogenesis. *Nature* 2012;481:463-468.
- 694 76 Rao RR, Long JZ, White JP, Svensson KJ, Lou J, Lokurkar I, Jedrychowski MP, Ruas JL, Wrann  
695 CD, Lo JC, Camera DM, Lachey J, Gygi S, Seehra J, Hawley JA, Spiegelman BM. Meteorin-like is

- 696 a hormone that regulates immune-adipose interactions to increase beige fat thermogenesis.  
697 Cell 2014;157:1279-1291.
- 698 77 Vaughan RA, Gannon NP, Mermier CM, Conn CA. Irisin, a unique non-inflammatory myokine  
699 in stimulating skeletal muscle metabolism. J Physiol Biochem 2015
- 700 78 Roberts LD, Bostrom P, O'Sullivan JF, Schinzel RT, Lewis GD, Dejam A, Lee YK, Palma MJ,  
701 Calhoun S, Georgiadi A, Chen MH, Ramachandran VS, Larson MG, Bouchard C, Rankinen T,  
702 Souza AL, Clish CB, Wang TJ, Estall JL, Soukas AA, Cowan CA, Spiegelman BM, Gerszten RE.  
703 Beta-aminoisobutyric acid induces browning of white fat and hepatic beta-oxidation and is  
704 inversely correlated with cardiometabolic risk factors. Cell Metab 2014;19:96-108.
- 705 79 Lee C, Zeng J, Drew BG, Sallam T, Martin-Montalvo A, Wan J, Kim SJ, Mehta H, Hevener AL, de  
706 Cabo R, Cohen P. The mitochondrial-derived peptide mots-c promotes metabolic  
707 homeostasis and reduces obesity and insulin resistance. Cell Metab 2015;21:443-454.
- 708 80 Craig DM, Ashcroft SP, Belew MY, Stocks B, Currell K, Baar K, Philp A. Utilizing small nutrient  
709 compounds as enhancers of exercise-induced mitochondrial biogenesis. Front Physiol  
710 2015;6:296.
- 711 81 Nogueira L, Ramirez-Sanchez I, Perkins GA, Murphy A, Taub PR, Ceballos G, Villarreal FJ,  
712 Hogan MC, Malek MH. (-)-epicatechin enhances fatigue resistance and oxidative capacity in  
713 mouse muscle. J Physiol 2011;589:4615-4631.
- 714 82 Borniquel S, Valle I, Cadenas S, Lamas S, Monsalve M. Nitric oxide regulates mitochondrial  
715 oxidative stress protection via the transcriptional coactivator pgc-1alpha. Faseb J  
716 2006;20:1889-1891.
- 717 83 Westerheide SD, Bosman JD, Mbadugha BN, Kawahara TL, Matsumoto G, Kim S, Gu W, Devlin  
718 JP, Silverman RB, Morimoto RI. Celastrols as inducers of the heat shock response and  
719 cytoprotection. J Biol Chem 2004;279:56053-56060.
- 720 84 Ma X, Xu L, Alberobello AT, Gavrilova O, Bagattin A, Skarulis M, Liu J, Finkel T, Mueller E.  
721 Celastrol protects against obesity and metabolic dysfunction through activation of a hsf1-  
722 pgc1alpha transcriptional axis. Cell Metab 2015;22:695-708.
- 723 85 Miller RA, Harrison DE, Astle CM, Baur JA, Boyd AR, de Cabo R, Fernandez E, Flurkey K, Javors  
724 MA, Nelson JF, Orihuela CJ, Pletcher S, Sharp ZD, Sinclair D, Starnes JW, Wilkinson JE, Nadon  
725 NL, Strong R. Rapamycin, but not resveratrol or simvastatin, extends life span of genetically  
726 heterogeneous mice. J Gerontol A Biol Sci Med Sci 2011;66:191-201.
- 727 86 Pearson KJ, Baur JA, Lewis KN, Peshkin L, Price NL, Labinsky N, Swindell WR, Kamara D,  
728 Minor RK, Perez E, Jamieson HA, Zhang Y, Dunn SR, Sharma K, Pleshko N, Woollett LA, Csiszar  
729 A, Ikeno Y, Le Couteur D, Elliott PJ, Becker KG, Navas P, Ingram DK, Wolf NS, Ungvari Z,  
730 Sinclair DA, de Cabo R. Resveratrol delays age-related deterioration and mimics  
731 transcriptional aspects of dietary restriction without extending life span. Cell Metab  
732 2008;8:157-168.
- 733 87 Novelle MG, Wahl D, Dieguez C, Bernier M, de Cabo R. Resveratrol supplementation: Where  
734 are we now and where should we go? Ageing Res Rev 2015;21:1-15.
- 735 88 Park EJ, Pezzuto JM. The pharmacology of resveratrol in animals and humans. Biochim  
736 Biophys Acta 2015;1852:1071-1113.
- 737 89 Tang PC, Ng YF, Ho S, Gyda M, Chan SW. Resveratrol and cardiovascular health--promising  
738 therapeutic or hopeless illusion? Pharmacol Res 2014;90:88-115.
- 739 90 Mouchiroud L, Houtkooper RH, Auwerx J. Nad(+) metabolism: A therapeutic target for age-  
740 related metabolic disease. Crit Rev Biochem Mol Biol 2013;48:397-408.
- 741 91 Shimabayashi M, Hall MN. Making new contacts: The mTOR network in metabolism and  
742 signalling crosstalk. Nat Rev Mol Cell Biol 2014;15:155-162.
- 743 92 Blattler SM, Cunningham JT, Verdeguer F, Chim H, Haas W, Liu H, Romanino K, Ruegg MA,  
744 Gygi SP, Shi Y, Puigserver P. Yin yang 1 deficiency in skeletal muscle protects against  
745 rapamycin-induced diabetic-like symptoms through activation of insulin/IGF signaling. Cell  
746 Metab 2012;15:505-517.
- 747 93 Martin-Montalvo A, Mercken EM, Mitchell SJ, Palacios HH, Mote PL, Scheibye-Knudsen M,  
748 Gomes AP, Ward TM, Minor RK, Blouin MJ, Schwab M, Pollak M, Zhang Y, Yu Y, Becker KG,

- 749                   Bohr VA, Ingram DK, Sinclair DA, Wolf NS, Spindler SR, Bernier M, de Cabo R. Metformin  
750                   improves healthspan and lifespan in mice. *Nat Commun* 2013;4:2192.
- 751           94       Check Hayden E. Anti-ageing pill pushed as bona fide drug. *Nature* 2015;522:265-266.
- 752           95       Grundlingh J, Dargan PI, El-Zanfaly M, Wood DM. 2,4-dinitrophenol (dnp): A weight loss agent  
753                   with significant acute toxicity and risk of death. *J Med Toxicol* 2011;7:205-212.
- 754           96       Kharitonenkova A, DiMarchi R. Fgf21 revolutions: Recent advances illuminating fgf21 biology  
755                   and medicinal properties. *Trends Endocrinol Metab* 2015;26:608-617.
- 756           97       Zhang Y, Xie Y, Berglund ED, Coate KC, He TT, Katafuchi T, Xiao G, Potthoff MJ, Wei W, Wan Y,  
757                   Yu RT, Evans RM, Kliewer SA, Mangelsdorf DJ. The starvation hormone, fibroblast growth  
758                   factor-21, extends lifespan in mice. *Elife* 2012;1:e00065.
- 759           98       Calvo JA, Daniels TG, Wang X, Paul A, Lin J, Spiegelman BM, Stevenson SC, Rangwala SM.  
760                   Muscle-specific expression of ppargamma coactivator-1alpha improves exercise performance  
761                   and increases peak oxygen uptake. *J Appl Physiol* 2008;104:1304-1312.
- 762           99       Choi CS, Befroy DE, Codella R, Kim S, Reznick RM, Hwang YJ, Liu ZX, Lee HY, Distefano A,  
763                   Samuel VT, Zhang D, Cline GW, Handschin C, Lin J, Petersen KF, Spiegelman BM, Shulman GI.  
764                   Paradoxical effects of increased expression of pgc-1alpha on muscle mitochondrial function  
765                   and insulin-stimulated muscle glucose metabolism. *Proc Natl Acad Sci U S A* 2008;105:19926-  
766                   19931.
- 767           100      Summermatter S, Baum O, Santos G, Hoppeler H, Handschin C. Peroxisome proliferator-  
768                   activated receptor {gamma} coactivator 1{alpha} (pgc-1{alpha}) promotes skeletal muscle  
769                   lipid refueling in vivo by activating de novo lipogenesis and the pentose phosphate pathway.  
770                   *J Biol Chem* 2010;285:32793-32800.
- 771           101      Dube JJ, Amati F, Stefanovic-Racic M, Toledo FG, Sauers SE, Goodpaster BH. Exercise-induced  
772                   alterations in intramyocellular lipids and insulin resistance: The athlete's paradox revisited.  
773                   *Am J Physiol Endocrinol Metab* 2008;294:E882-888.
- 774           102      Summermatter S, Shui G, Maag D, Santos G, Wenk MR, Handschin C. Pgc-1alpha improves  
775                   glucose homeostasis in skeletal muscle in an activity-dependent manner. *Diabetes*  
776                   2013;62:85-95.
- 777           103      Summermatter S, Handschin C. Pgc-1alpha and exercise in the control of body weight. *Int J  
778                   Obes (Lond)* 2012;36:1428-1435.
- 779           104      Handschin C. The biology of pgc-1alpha and its therapeutic potential. *Trends Pharmacol Sci*  
780                   2009;30:322-329.

781

782

783 **Figure legends**

784

785 **Figure 1. Common and distinct effects of exercise and caloric restriction.** Even though exercise and  
786 caloric restriction affect energy intake (at least in some individuals) and expenditure in a  
787 diametrically opposite manner, the shared regulation of a number of phenotypic changes in skeletal  
788 muscle and potentially other tissues could underlie the similar health benefits of both interventions.  
789 Importantly however, other effects, e.g. on muscle and cardiovascular function as well as body  
790 weight, are predominantly observed after exercise and caloric restriction, respectively.

791

792 **Figure 2. Molecular signaling of exercise and caloric restriction “mimetics” centered on PGC-1 $\alpha$ .**  
793 Proposed mechanisms of action of several exercise and caloric restriction “mimetics” are depicted. \*  
794 indicates coactivation of the respective transcription factors by PGC-1 $\alpha$ . See text for details and  
795 abbreviations.

796

797

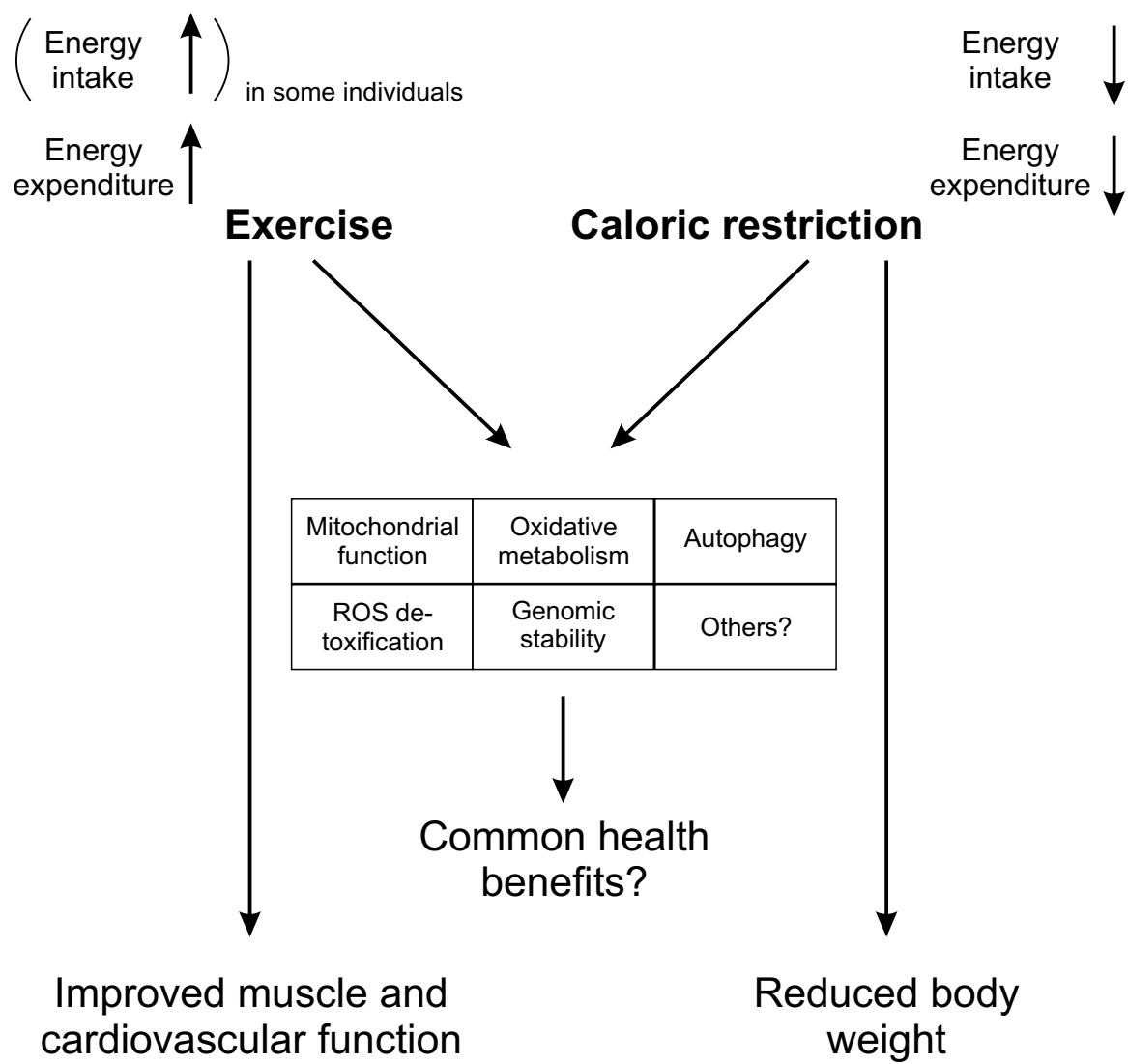


Figure 1

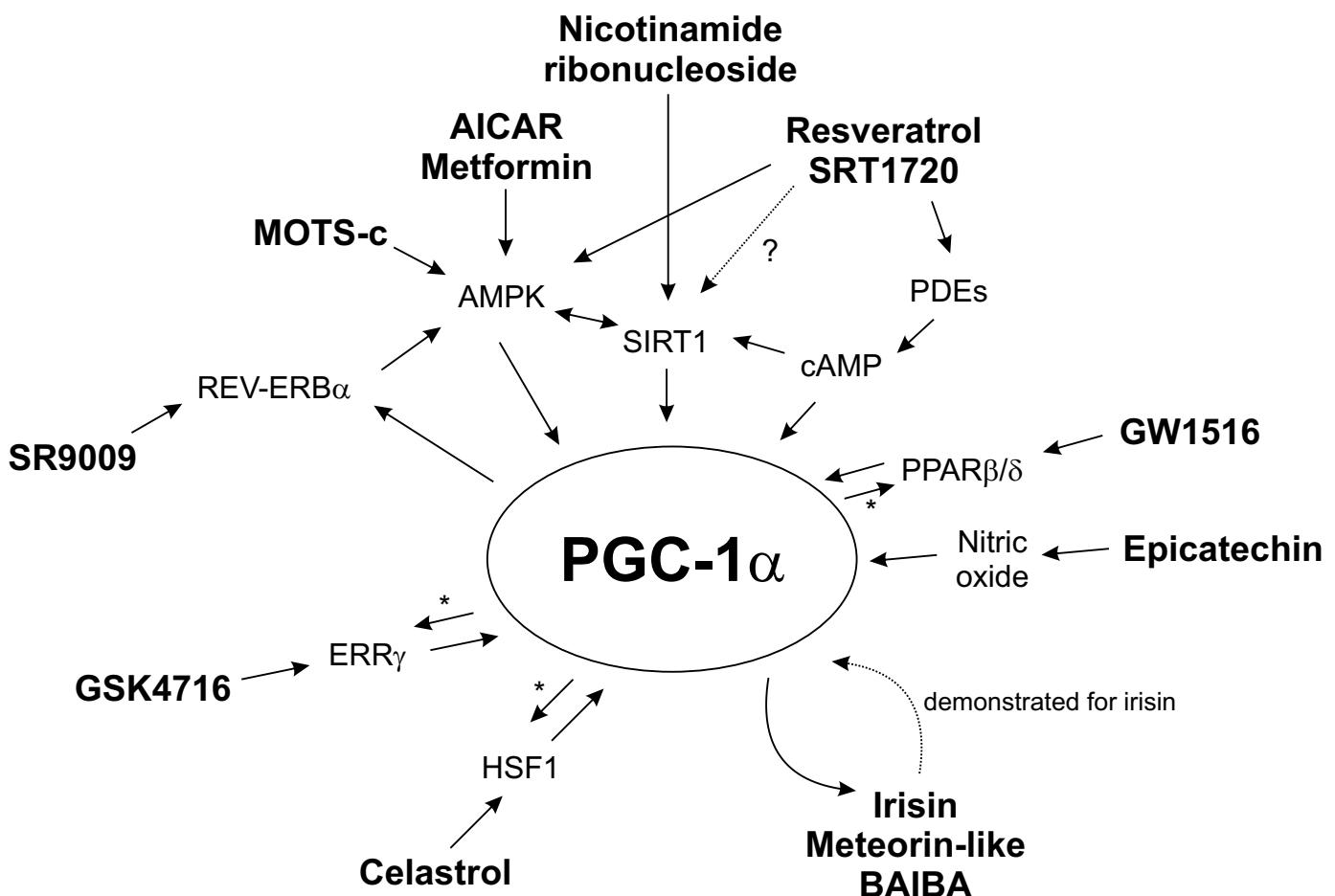


Figure 2

